

Reduction of phenolics in faba bean meal using recombinantly produced and purified *Bacillus ligniniphilus* catechol 2,3-dioxygenase

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Additional file Figures

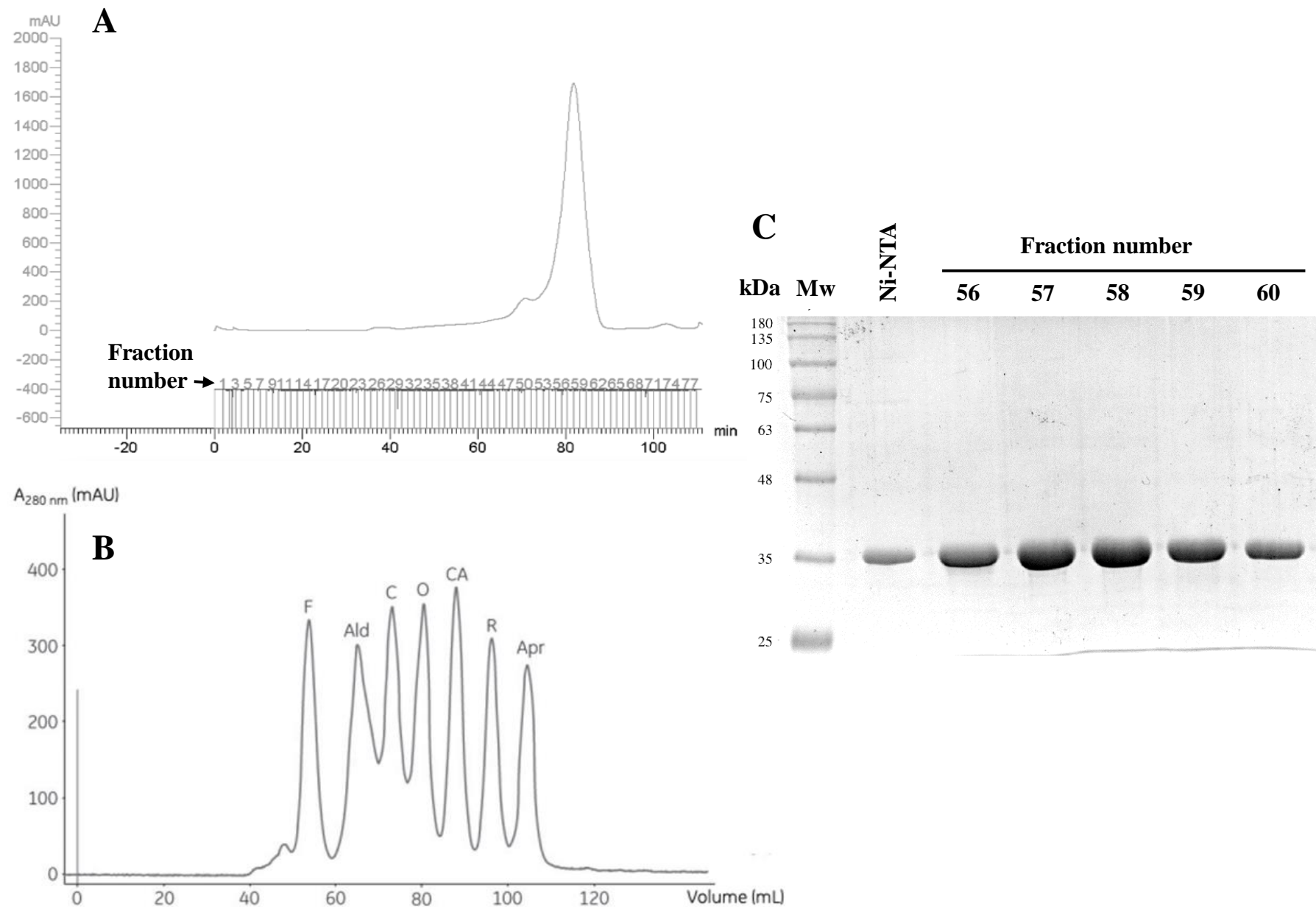


Figure S1:
Purification of BLC23O by size-exclusion chromatography. **A)** Size-exclusion chromatography (SEC) was performed with a HiLoad 16/600 Superdex 200 column attached to an FPLC system and run at a flowrate of 1 mL/min. **A)** The elution profile (OD_{280nm}) with the ladder at the bottom of the graph representing the fractions collected during the run. **B)** Elution profile for standard proteins to calibrate the SEC column where F is ferritin, Ald is aldolase, C is conalbumin, O is ovalbumin (45kDa), CA is carbonic anhydrase (29.2 kDa), R is RNase A, and Apr is aprotinin (Cytvio). **C)** A 12% acrylamide SDS-PAGE analysis of select fractions from the BLC23O SEC purification..

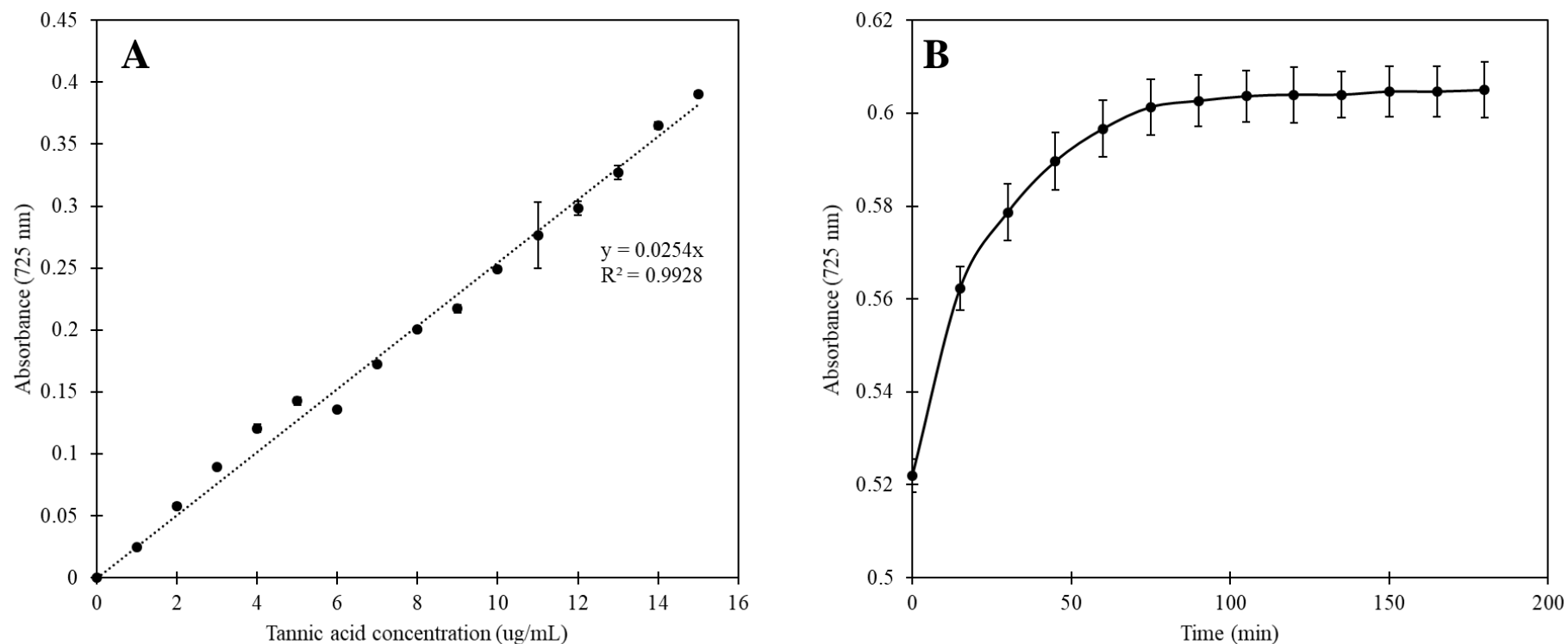


Figure S2: Folin-Ciocalteu assay development. **A)** Calibration curve for determination of total phenolic content. Folin-Ciocalteu reagent was added to known concentrations of tannic acid ranging from 0 to 30 ug/mL and left to react for 45 min. Absorbances were then measured at 725 nm. The data represents the mean and standard deviation (n=3). A linear fit of the data with an equation of $y=0.0127x$ and a R^2 value of 0.9981 are shown. **B)** Evaluation of Folin-Ciocalteu reagent kinetics. Reaction solutions with volumes of 250 μ L were prepared with 0.125 N Folin-Ciocalteu reagent, 0.125 mg/mL sodium carbonate, and 14 μ g/mL tannic acid. The absorbance at 725 nm was monitored for 180 min in 15 min intervals. The data represents the mean and standard deviation (n=3).

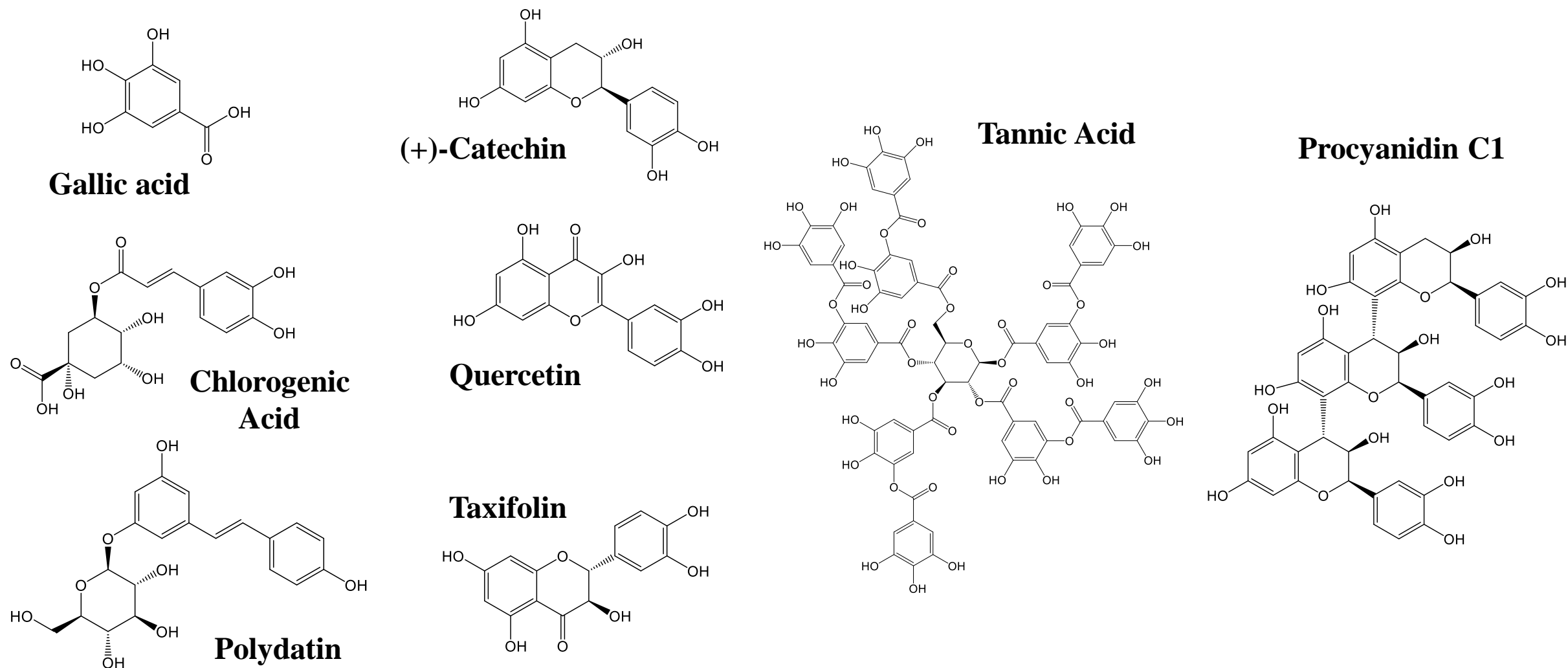


Figure S3: Representative images of commercially available phenolic compounds tested as substrates for BLC23O.

